

CHEMICAL CONSTITUENTS AND MEDICAL FUNCTION OF LEAVES OF *DIOSPYROS LOTUS* L.

by

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The potential use of leaves of Diospyros lotus L. was analyzed by using FT-IR, GC-MS and MTT through organic solvent extracts. The results showed that the leaves extracts of Diospyros lotus L. contains many kinds of bioactive organic ingredients with antibacterial, antitumor, and anti-inflammatory activities. Meanwhile the extracts have significant effect on HEPG2, SGC-7091, and K562. Therefore, it provides a scientific foundation for comprehensive application prospects in the fields of biomedicine, food, skin care products and other fields and turning waste into treasure.

Key words: *Diospyros lotus* L., FI-RT, GC-MS, MTT

Introduction

Plants are the richest sources which included bioactive compounds. All kinds of residual product of plants have different biological activities and play a very important role for maintaining human health and treatment of disease [1-5]. So many plants were used as traditional Chinese Medicine and have many unexpected effects that western medicine cannot achieved [6-8]. It is well known that *Diospyros lotus* L. is one of the plants with specific selectivity [9]. *Diospyros lotus* L. is a species of tree in the genus *Diospyros* and the family *Ebenaceae* and widely distributed in China. It has been cultivated in several countries for its edible fruits and used as traditional medicines for various medicinal purposes [10-17]. *Diospyros lotus* L. has more economic importance with yielding edible fruits and valuable timbers. Its timber was still famous and precious for furniture and wood products. Its leaves are approved as Chinese Medicine for the treatment of stroke and apoplexy syndrome in China or used as a hypotensive drug in Japan [18]. Furthermore, it has been used as herb tea and the treatment of hypertension in patients for long time in Korea [19-21]. However, many researches focused on the fruits and timber, little studies have been done for comprehensive use of the persimmon leaves [22-24]. The objective of the present study was to investigate the constituents of the leaves extracts of *Diospyros lotus* L. by FT-IR, GC-MS, and MTT analysis. Then biological activities ingredients

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were identified and prospect of resource utilization was prospected [25-27]. It is helpful for turning leaves waste into treasure.

Material and methods

Experimental materials

Samples were collected from Xixia forest area in Henan Province. Samples were processed from fresh material into powder and oven-dried to absolute dryness at 60 °C. Three kinds of extracts were named as L1, L2 and L3 samples which were extracted by ethanol, ethanol/benzene (1:2) and methanol respectively [28, 29].

Experimental methods

The FT-IR analysis

The FT-IR spectrum of the sample was obtained on a FT-IR spectrophotometer (IR100) using a KBr disk containing 1.00% finely ground samples. The scanning of each extracts was collected at a spectral resolution of 4 cm⁻¹ and the spectral range was 500-4000 cm⁻¹ [30].

The GC-MS analysis

The three extracts were analyzed by using a gas chromatography-mass spectrometer (Agilent GC-MS 7890B 5977A). Column HP-5MS (30 m × 250 μm × 0.25 μm). Elastic quartz capillary column, the carrier gas used for high purity helium, flow rate of 1 mL/min. The split ratio is 20:1. The temperature program of the GC starts at 50 °C, rises to 250 °C at a rate of 8 °C/min, and then rises to 300 °C at a rate of 5 °C/min. The MS program scan mass range of 30-600 amu, ionization voltage of 70 eV, ionization current of 150 μA electron ionization (EI). The ion source and the quadrupole temperature were set at 230 °C and 150 °C, respectively.

The MTT analysis

Cells are treated with an isopropanol-10% Triton X-100 solution, collected the solubilized formazan and transferred triplicate aliquots to a 96-well standard plate (NUNC). Formazan production was measured by spectrophotometry using the SOFTmax Pro microplate reader (Molecular Devices) equipped with the SOFTmax Pro software package (v.4.7).

Absorbance was collected at λ = 570 nm, subtracting the background measured at λ = 690 nm [31].

Results and discussion

Analysis of FT-IR

As the fig. 1 showed that the infrared spectra of three kinds of extracts of *Diospyros lotus* L. was similar. The O-H stretching vibration are at 3361 cm⁻¹, 3366 cm⁻¹ and 3357 cm⁻¹. The C-H and -CH₂- stretching vibration are at 2925 cm⁻¹ and 2852 cm⁻¹, respectively. Conjugated C=O stretching at 1692 cm⁻¹ and aromatic CH stretching at 1650-1430 cm⁻¹. 1613 cm⁻¹ and 1515

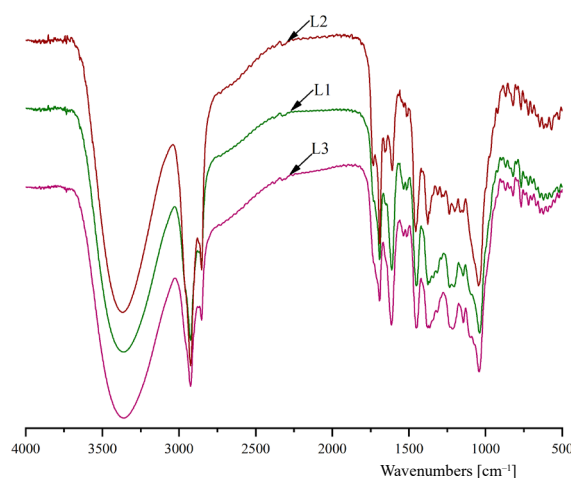


Figure 1. The FT-IR spectra of samples L1, L2, and L3

cm^{-1} is the aromatic carbon skeleton vibration. The absorption peaks of leaves extracts of the *Diospyros lotus* L. are mainly concentrated in the wave segments of $3700\text{--}3000\text{ cm}^{-1}$, $3000\text{--}2800\text{ cm}^{-1}$, and $1690\text{--}970\text{ cm}^{-1}$. The main chemical components are alcohols, acids, phenols and aromatic compounds [32-36]. The three absorption peaks have slight difference in infrared, which indicated that the main chemical components are basically identical.

Analysis of GC-MS

The total ion chromatograms of three kinds of extractives of ethanol, ethanol/benzene and methanol were shown in figs. 2-4, which were analyzed by GC-MS.

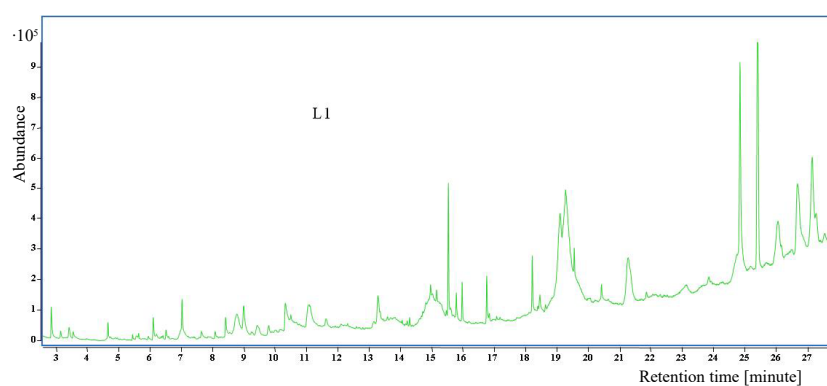


Figure 2. Total ion chromatograms of L1

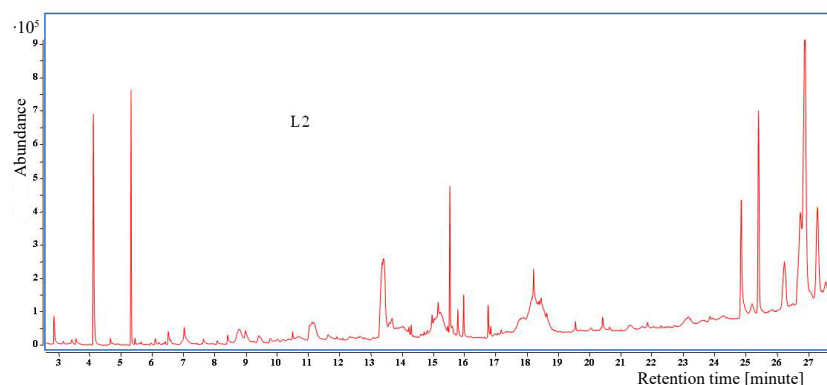


Figure 3. Total ion chromatograms of L2

According to the results of GC-MS analysis, 60 peaks were detected in L1, and 34 chemical constituents were identified. The components are: Glutinol(20.5%), 9-Octadecenamide, (Z)- (10.14%), Squalene (7.71%), 2H-3,9a-Methano-1-benzoxepin, octahydro-2,2,5a,9-tetramethyl-, [3R-(3.alpha.,5a.alpha.,9.alpha.,9a.alpha.)]- (7.53%), Melezitose (7.04%), Lupeol (6.72%), .gamma.-Sitosterol(6.42%), Vitamin E(6.22%), 3-n-Butylthiolane (3.96%), 2-Hexadecen-1-ol, 3,7,11,15-tetramethyl-, acetate, [R-[R*,R*-(E)]]- (3.64%), Lactose (2.97%), Ethylene glycol butyl ether, trimethylsilyl ether (2.14%), .beta.-Amyrin (1.94%), 2-Butyloxycarbonyloxy-1,1,10-trimethyl-6,9-epidioxydecalin (1.58%), 1,2,3-Benzenetriol (1.53%), 2,2-Dimethyl-6-methylene-1-[3,5-dihydroxy-1-pentenyl]cyclohexan-1-perhydropol (1.43%), n-Hexadecanoic acid (1.14%), (2R,3R,4aR,5S,8aS)-2-Hydroxy-4a,5-dimethyl-

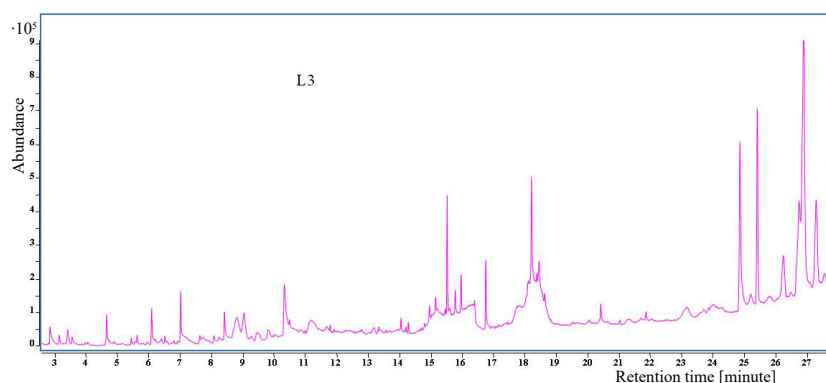


Figure 4. Total ion chromatograms of L3

3-(prop-1-en-2-yl)octahydronaphthalen-1(2H)-one (0.94%), N,N-Dimethyl-O-(1-methyl-butyl)-hydroxylamine (0.91%), Decanoic acid, 3-hydroxy-, methyl ester (0.68%), Thymine (0.66%), 9,12-Octadecadienoic acid (Z,Z)- (0.64%), Cyclohexanol, 3-ethenyl-3-methyl-2-(1-methylethenyl)-6-(1-methylethyl)-, [1R-(1.alpha.,2.alpha.,3.beta.,6.alpha.)]- (0.63%), Bi-1-cyclohexen-1-yl, 3,3,3',3',5,5,5',5'-octamethyl- (0.47%), Undec-10-ynoic acid, propyl ester (0.45%), 2,4-Dimethyl-5-methylthiopent-4-en-2-ol (0.41%), 5-Pregnene, 3-acetoxy-20-[(N-acetyl-4-methylpyrrolidin-2-yl)carbonyl]- (0.29%), p-Toluidine, N-methyl-N-nitroso- (0.24%), Uridine, 5-methoxy- (0.23%), Phen-1,4-diol, 2,3-dimethyl-5-trifluoromethyl- (0.22%), 2-Hexenoic acid, 5-hydroxy-3,4,4-trimethyl-, (E)- (0.17%), 9-Decenoic acid (0.17%), Silane, 1,3-heptadiynyltrimethyl- (0.14%), .beta.-D-Glucopyranose, 1-thio-,1-[N-hydroxy-5-(methylthio)pentanimidate] (0.13%).

According to the results of GC-MS analysis, 45 peaks were detected in L2, and 32 chemical constituents were identified. The components are: 1,4-Dimethyl-8-isopropylidenetri cyclo[5.3.0.0(4,10)]decane (19.17%), .beta.-Amyrin (10.64%), Lupeol (8.88%), .beta.-D-Glucopyranoside, methyl (8.42%), Lup-20(29)-en-3-one (7.88%), Squalene (5.58%), Lactose (5.12%), Triethylamine (4.54%), 9-Octadecenamamide, (Z)- (4.21%), Heptane, 1,1'-oxybis- (3.61%), Androst-1-en-3-one, 4,4-dimethyl-, (5.alpha.)- (3.05%), .beta.-Amyrone (2.98%), .alpha.-d-6,3-Furanose, methyl-.beta.-d-glucohexodialdo-1,4-furanoside (2.68%), Neophytadiene (2.14%), (2R,3R,4aR,5S,8aS)-2-Hydroxy-4a,5-dimethyl-3-(prop-1-en-2-yl)octahydronaphthalen-1(2H)-one (1.53%), Ethylene glycol butyl ether, trimethylsilyl ether (1.43%), .beta.-D-Glucopyranose, 1-thio-,1-[N-hydroxy-5-(methylthio)pentanimidate] (0.96%), 2H-3,9a-Methano-1-benzoxepin, octahydro-2,2,5a,9-tetramethyl-, [3R-(3.alpha.,5a.alpha.,9.alpha.,9a.alpha.)]- (0.85%), 2-Hexadecen-1-ol, 3,7,11,15-tetramethyl-, acetate, [R-[R*,R*-(E)]]- (0.72%), N,N-Dimethyl-O-(1-methyl-butyl)-hydroxylamine (0.71%), n-Hexadecanoic acid (0.59%), .alpha.-Amyrin (0.56%), Melezitose (0.53%), Vitamin E (0.48%), d-Glycero-l-glucoc-heptose (0.48%), Galactoseptanoside, methyl 2,3,4,5-tetra-O-methyl-, .alpha.-D- (0.46%), Inositol, 1-deoxy- (0.41%), 17-Octadecynoic acid (0.39%), Decanoic acid, 3-hydroxy-, methyl ester (0.31%), Silane, 1,3-heptadiynyltrimethyl- (0.28%), 2,2-Dimethyl-6-methylene-1-[3,5-dihydroxy-1-pentenyl]cyclohexan-1-perhydrol (0.22%), Mannosamine (0.20%).

According to the results of GC-MS analysis, 44 peaks were detected in L3, and 29 chemical constituents were identified. The components are: 1H-3a,7-Methanoazulen-6-ol, octahydro-3,6,8,8-tetramethyl-, acetate, [3R-(3.alpha.,3a.beta.,6.alpha.,7.beta.,8a.alpha.)]- (20.63%), .beta.-Amyrin (12.09%), Lup-20(29)-en-3-one (8.77%), 13-Docosenamamide, (Z)- (6.72%), Squalene (5.84%), (2R,3R,4aR,5S,8aS)-2-Hydroxy-4a,5-dimethyl-3-(prop-1-en-

2-yl)octahydronaphthalen-1(2H)-one (5.78%), Lupeol (4.15%), 3-n-Butylthiolane (4.14%), Melezitose (3.66%), .beta.-Amyrone (3.61%), D-Galactose (3.30%), 1,2,3-Benzenetriol (3.15%), Farnesyl bromide (2.96%), Lactose (2.52%), 2-Hexadecen-1-ol, 3,7,11,15-tetramethyl-, acetate, [R-[R*,R*-(E)]]- (2.11%), n-Hexadecanoic acid (1.54%), Decanoic acid, 3-hydroxy-, methyl ester (1.25%), 2,2-Dimethyl-6-methylene-1-[3,5-dihydroxy-1-pentenyl] cyclohexan-1-perhydrol (1.19%), D-chiro-Inositol, 3-O-(2-amino-4-((carboxyiminomethyl) amino)-2,3,4,6-tetraoxy-.alpha.-D-arabino-hexopyranosyl)- (1.15%), Thymine (1.04%), .alpha.-Amyrin (0.96%), 2,4-Dihydroxy-2,5-dimethyl-3(2H)-furan-3-one (0.78%), N,N-Dimethyl-O-(1-methyl-butyl)-hydroxylamine (0.76%), Epi-Inositol (0.57%), Thiophene, tetrahydro-2-methyl- (0.55%), d-Glucosamine (0.30%), d-Glycero-l-gluco-heptose (0.28%), Maltose (0.23%), [1,1'-Bicyclohexyl]-2-one (0.23%).

The identified compounds can be classified into alcohols, acids, phenols, squalene, and so on. The extracts contain the Lup-20(29)-en-3-one and Lupeol, which are the active ingredient of tumor suppression and prevention and can reduced risk of cancer, especially breast and prostate cancer [37-39]. The .beta.-Amyrin can inhibit the synthesis of cholesterol and triglycerides and prevent effectively obesity, atherosclerosis and type 2 diabetes. Squalene is the active ingredient antibacterial anti-inflammatory and has the effect on hypertension hepatocirrhosis, diabetes, brochitis and other diseases. Meanwhile their content was higher in the three extracts. Lactose is widely used in making baby food, candy and other food industries. The n-Hexadecanoic acid is important raw material of soap, candles, lubricants, softeners and synthetic detergents. Triethylamine is a key intermediate in the synthesis of AZT, DDT and related drugs, which was extracted only by ethanol/benzene. Glutanol is a compound with known anti-inflammatory activity which was only extracted by ethanol and higher content. Vitamin E is used for the prevention of habitual abortion and threatened abortion caused by vitamin E deficiency as well as for protecting cells and tissues from lipoperoxidative damage induced by free radicals [40]. The other compounds can be used in the industry of food, chemical product, medicine, and the like. The results also showed that the representative compounds contents with more than 3% accounted for more 80% of the total extracted components respectively. Meanwhile it was found that 11 components of the three extracts were the same, and all of them were more than one third of the total extract content by comparison.

Analysis of MTT

Sample L1 (extracted by ethanol) was selected for MTT experiment [41-46]. The results showed that the L1 has a strong anticancer effect on HEGP2, SGC-7091, and K562. The killing rate was 44.97%, 40.47%, and 28.29% respectively. The antitumor effect on HEGP2 cells was the most obvious. Some ingredients in the leaves of *Diospyros lotus* L. can effectively prevent and cure cancer, which can be used for Chinese medical industry.

Conclusion

The main chemical constituents of the leaves extracts of the *Diospyros lotus* L. are alcohols, acids, phenols, ethers and aromatic compounds. The biological activities ingredients have antibacterial, anti-inflammatory, antitumor and antioxidant, which can be used in medicine, food, skin care products. Furthermore, the leaves extracts could be used as Chinese traditional medicine directly.

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References

- [1] Cho, B. O., *et al.*, Ameliorative Effects of, *Diospyros Lotus*, Leaf Extract Against UVB-Induced Skin Damage In Balb/C Mice, *Biomedicine & Pharmacotherapy*, 95 (2017), Nov., pp. 264-274
- [2] Uddin, G., *et al.*, Anti-Nociceptive, Anti-Inflammatory and Sedative Activities of the Extracts and Chemical Constituents of *Diospyros Lotus* L., *Phytomedicine*, 21 (2014), 7, pp. 954-959
- [3] Sa, Y. S., *et al.*, The Anticoagulant Fraction from the Leaves of *Diospyros Kaki* L. Has an Antithrombotic Activity, *Archives of Pharmacal Research*, 28 (2005), 6, pp. 667-674
- [4] Rauf, A., *et al.*, In Vivo and in Silico Sedative-Hypnotic Like Activity of 7-Methyljuglone Isolated from *Diospyros Lotus* L., *Biomedicine & Pharmacotherapy*, 87 (2017), Mar., pp. 678-682
- [5] Adams, J. D., Wang, X., Control of Pain with Topical Plant Medicines, *Asian Pacific Journal of Tropical Biomedicine*, 5 (2015), 4, pp. 268-273
- [6] Loizzo, M. R., *et al.*, Antioxidant and Antiproliferative Activity of *Diospyros Lotus* L. Extract and Isolated Compounds, *Plant Foods for Human Nutrition*, 64 (2009), 4, pp. 264-270
- [7] Zadbakht, M., *et al.*, *Diospyros Lotus* L. Fruit Extract Protects G6PD-Deficient Erythrocytes from Hemolytic Injury in Vitro and in Vivo: Prevention of Favism Disorder, *Planta Medica*, 15 (2011), 11, pp. 1270-1281
- [8] Faiz, O., Baltas, N., Polyphenol Oxidase Properties, Anti-Urease, and Anti-Acetylcholinesterase Activity of *Diospyros Lotus* L. (Plum Persimmon), *International Journal of Food Properties*, 20 (2017), 5, pp. 1186-1196
- [9] Nabavia, S. M., *et al.*, In Vitro Antioxidant and Free Radical Scavenging Activity of *Diospyros Lotus* and *Pyrus Boissieriana* Growing in Iran, *Pharmacognosy Magazine*, 4 (2009), 18, pp. 122-126
- [10] Arnal, L., Del Rio, M. A., Effect of Cold Storage and Removal Astringency on Quality of Persimmon Fruit (*Diospyros Kaki*, L.) Cv. Rojo Brillante, *Food Science & Technology International*, 10 (2004), 3, pp. 179-185
- [11] Hu, D., *et al.*, Phylogenetic Analysis in Some *Diospyros* Spp. (Ebenaceae) and Japanese Persimmon Using Chloroplast Dna Pcr-Rflp Markers, *Scientia Horticulturae*, 117 (2008), 2, pp. 32-38
- [12] Yang, Y., *et al.*, Genetic Diversity and Taxonomic Studies of Date Plum (*Diospyros Lotus* L.) Using Morphological Traits and Scot Markers, *Biochemical Systematics and Ecology*, 61 (2015), Aug., pp. 253-259
- [13] Rauf, A., *et al.*, Anti-Tumour-Promoting and Thermal-Induced Protein Denaturation Inhibitory Activities of B-Sitosterol and Lupeol Isolated From *Diospyros Lotus* L., *Natural Product Research*, 30 (2015), 10, pp. 1205-1207
- [14] Rauf, A., *et al.*, *Diospyros*, an under-Utilized, Multi-Purpose Plant Genus: A Review, *Biomedicine & Pharmacotherapy*, 91 (2017), July, pp. 714-730
- [15] Adebayo, S. A., *et al.*, First Isolation of Glutinol and a Bioactive Fraction with Good Antiinflammatory Activity from N-Hexane Fraction of *Peltophorum Africanum* Leaf, *Asian Pacific Journal of Tropical Medicine*, 10 (2017), 1, pp. 42-46
- [16] Chen, J. T., *et al.*, Molecules and Functions of Rosewood: *Diospyros Celebica*, *Arabian Journal of Chemistry*, 11 (2017), 6, pp. 756-762
- [17] Glew, R. H., *et al.*, Changes in Sugars, Acids and Fatty Acids in Naturally Parthenocarpic Date Plum Persimmon (*Diospyros Lotus* L.) Fruit During Maturation and Ripening, *European Food Research and Technology*, 221 (2005), 1-2, pp. 113-118
- [18] Fan, J. P., He, C. H., Simultaneous Quantification of Three Major Bioactive Triterpene Acids in the Leaves of *Diospyros Kaki* by High-Performance Liquid Chromatography Method, *J Pharm Biomed Anal*, 41 (2006), 3, pp. 950-956
- [19] Lee, M. S., Sohn, C. B., Anti-Diabetic Activities of Ethanol Extracts from Persimmon Leaves, *Journal of the Korean Society for Applied Biological Chemistry*, 52 (2009), 1, pp. 96-97
- [20] Sun, L., *et al.*, Evaluation to the Antioxidant Activity of Total Flavonoids Extract from Persimmon (*Diospyros Kaki* L.) Leaves, *Food & Chemical Toxicology*, 49 (2011), 10, pp. 2689-2696
- [21] Lee, S. G., *et al.*, Immunostimulatory Polysaccharide Isolated from the Leaves of *Diospyros Kaki*, Thumb Modulate Macrophage Via, TLR2, *International Journal of Biological Macromolecules*, 79 (2015), Aug., pp. 971-982

- [22] Said, A., *et al.*, Pharmaco-Chemical Studies on the Aqueous Methanolic Extract of *Diospyros Lotus* Leaves, *Research Journal of Phytochemistry*, 3 (2009), 1, pp. 1-12
- [23] Kiaei, M., Bakhshi, R., Short Communication Radial Variations of Wood Different Properties in *Diospyros Lotus*, *Forest Systems*, 23 (2014), 1, pp. 171-177
- [24] Rauf, A., *et al.*, In Vivo Sedative and Muscle Relaxants Activity of *Diospyros Lotus* L, *Asian Pacific Journal of Tropical Biomedicine*, 5 (2015), 4, pp. 277-280
- [25] Rout, S. P., Kar, D. M., Identification of Chemical Compounds Present in Different Fractions of *Annona Reticulata* L. Leaf by Using GC-MS, *Natural Product Research*, 28 (2014), 20, pp. 1786-1788
- [26] Fernandez-Pousa, C.R., Perfect Phase-Coded Pulse Trains Generated by Talbot Effect, *Applied Mathematics & Nonlinear Sciences*, 3 (2018), 1, pp. 23-32
- [27] Loksha, V., *et al.*, Operations of Nanostructures Via Sdd, Abc4 and Ga5 Indices, *Applied Mathematics & Nonlinear Sciences*, 2 (2017), 1, pp. 173-180
- [28] Wanxi, P., *et al.*, Immune Effects of Extractives on Bamboo Biomass Self-Plasticization, *Pakistan Journal of Pharmaceutical Sciences*, 27 (2014), 4, pp. 991-999
- [29] Li, C., *et al.*, Structural Properties and Copolycondensation Mechanism of Valonea Tannin-Modified Phenol-Formaldehyde Resin, *Journal of Polymers and the Environment*, 26 (2018), 3, pp. 1297-1309
- [30] Melin, T., Ft-ir Analysis of BSA Fouled on Ultrafiltration and Microfiltration Membranes, *Journal of Membrane Science*, 192 (2001), 1, pp. 201-207
- [31] Angius, F., Floris, A., Liposomes and MTT Cell Viability Assay: An Incompatible Affa, *Toxicology in Vitro*, 29 (2015), 2, pp. 314-319
- [32] Iwaki, L. K., Dlott, D. D., Ultrafast Vibrational Energy Redistribution Within C-H and O-H Stretching Modes of Liquid Methanol, *Chemical Physics Letters*, 321 (2000), 5-6, pp. 419-425
- [33] Botha, A., Strydom, C. A., DTA and FT-IR Analysis of the Rehydration of Basic Magnesium Carbonate, *Journal of Thermal Analysis and Calorimetry*, 71 (2003), 3, pp. 987-996
- [34] Yao, X., *et al.*, Distinction of Eight Lycium Species by Fourier-Transform Infrared Spectroscopy and Two-Dimensional Correlation IR Spectroscopy, *Journal of Molecular Structure*, 974 (2010), 1-3, pp. 161-164
- [35] Xu, G., *et al.*, FTIR and XPS Analysis of the Changes in Bamboo Chemical Structure Decayed by White-Rot and Brown-Rot Fungi, *Applied Surface Science*, 280 (2013), Sept., pp. 799-805
- [36] Tomak, E. D., *et al.*, An FT-IR Study of the Changes in Chemical Composition of Bamboo Degraded by Brown-Rot Fungi, *International Biodeterioration & Biodegradation*, 85 (2013), Nov., pp. 131-138
- [37] Saleem, M., Lupeol, a Novel Anti-Inflammatory and Anti-Cancer Dietary Triterpene, *Cancer Letters*, 285 (2009), 2, pp. 109-115
- [38] Saleem, M., Lupeol, a Fruit and Vegetable Based Triterpene, Induces Apoptotic Death of Human Pancreatic Adenocarcinoma Cells Via Inhibition of Ras Signaling Pathway, *Carcinogenesis*, 26 (2005), 11, pp. 1956-1964
- [39] Siddique, H. R., Saleem, M., Beneficial Health Effects of Lupeol Triterpene: A Review of Preclinical Studies, *Life Sciences*, 88 (2011), 7-8, pp. 285-293
- [40] Gao, H., *et al.*, Antioxidant Activities and Phenolic Compounds of Date Plum Persimmon (*Diospyros Lotus* L.) Fruits, *Journal of Food Science and Technology*, 51 (2014), 5, pp. 950-956
- [41] Berridge, M. V., *et al.*, Tetrazolium Dyes as Tools in Cell Biology: New Insights Into Their Cellular Reduction, *Biotechnology Annual Review*, 11 (2005), Feb., pp. 127-152
- [42] Ahmad, S., *et al.*, Cholesterol Interferes with the MTT Assay in Human Epithelial-Like (A549) and Endothelial (HLMVE and HCAE) Cells, *International Journal of Toxicology*, 25 (2006), 1, pp. 17-23
- [43] Meerloo, J. V., *et al.*, Cell Sensitivity Assays: The MTT Assay, *Methods Mol Biol*, 731 (2011), Mar., pp. 237-245
- [44] Shen, J., *et al.*, Antitumor Activity of Cobrotoxin in Human Lung Adenocarcinoma A549 Cells and Following Transplantation in Nude Mice, *Oncology Letters*, 8 (2014), 5, pp. 1961-1965
- [45] Grela, E., *et al.*, Current Methodology of MTT Assay in Bacteria – A Review, *Acta Histochemica*, 120 (2018), 4, pp. 303-311
- [46] Jiang, S. C., *et al.*, Molecules and Functions of Rosewood: *Dalbergia Stevenson*, *Arabian Journal of Chemistry*, 11 (2017), 6, pp. 782-792